A xeno-free slow-freezing cryopreservation medium for primate ES/iPS cells

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Abstract

Cryopreservation of primate ES/iPS cells is severe and difficult compared to murine ones. It can be described that vitrification method might be adequate for primate ES/iPS cells at this moment, however it needs special skills and has to avoid dry ice transportation. To address these problems, we developed a new xeno-free freezing medium for primate ES/iPS cells using slow-freezing, and evaluated its recovery and dry ice transportation.

Introduction

Materials & method

A flowchart of cryopreservation, transportation and thawing for primate ES/iPS cells

A) Slow freezing



B) Vitrification



The recovery of primate ES/iPS cells after cryopreservation with the conventional slow-freezing is quite low .Vitrification is not an easy option because the steps required are more complicated than those of slow-freezing. In addition, frozen cells using vitrification have to be avoided dry ice transportation. Therefore more efficient freezing method or freezing medium are needed.

Common marmoset ES cell line, CMEM40 and human iPS cell line, ATCC-HYR0103 were used to evaluate the efficiency of slow-freezing or vitrification for cryopreservation of primate ES/iPS cells. The ES cells were dissociated into several small clumps and frozen in our freezing medium or 10% DMSO/medium by slow-freezing or in a vitrification solution by quick-freezing. After thawing, cells were cultured for 4 days. The number of alkaline phosphatase-positive colonies was counted as recovery points.

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The ES cells were frozen as described above. The frozen cells were transferred to a box filled with dry ice and stored for 24 hours. After thawing, ES cells were cultured for 4 days. The recovery was estimated as described above.

10% DMSO/medium,	Our freezing medium
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-80°C	LN2		Dry ice)	
day1	2	3	4	5	
Vitrification solution					
LN2			Dry ice)	
day1	2	3	4	5	

The iPS cells were cultured in the presence or absence of Y-27632(ROCK inhibitor) before freezing. 1 hour later, colonies were dissociated into single cells and cells were frozen in our freezing medium using slow-freezing. After thawing, 20,000 cells were seeded and

Result

cultured in the presence or absence of Y-27632. 18 hours later, culture medium was changed in the absence of Y-27632 and cells were cultured for 8 days. The recovery was estimated as described above.



Figure 1. The recovery of monkey ES cells after cryopreservation and thawing

Figure 2. The loss rate of colonies after dry ice storage

Figure 3. The recovery of human iPS cells in the presence of the Y-27632 (ROCK inhibitor)

The ES cells were cryopreserved in 10% DMSO/ medium(A) or vitrification solution(B) or our freezing medium(C). The recovery efficiency was estimated by counting the number of alkaline phosphatase-positive colonies at 4 days after thawing. Results are expressed as means \pm SEM for n=3.*P<0.05, **P<0.01 The ES cells were cryopreserved in 10% DMSO/medium(A) or vitrification solution(B) or our freezing medium(C) then stored in a box filled with dry ice for 24 hours. The loss rates of colony were calculated by comparing the number of colonies after dry ice storage/liquid nitrogen storage at 4 days after thawing. Results are expressed as means \pm SEM for n=3.

The iPS cells were cultured in the presence or absence of Y-27632 before freezing.1 hour later, colonies were dissociated into single cells and cells were frozen in our freezing medium using slow-freezing. Cells were thawed and 20,000 cells were seeded in the presence or absence of Y-27632. The recovery was estimated by counting the number of alkaline phosphatase-positive colonies at 8 days after thawing. Results are expressed as means \pm SEM for n=3. *P<0.001

Conclusion

We have developed an efficient freezing medium for primate ES/iPS cells.
Cells frozen with our medium can be transported using a container filled with dry ice.
Our medium is quite useful as xeno-free freezing medium especially in medical application.