

## Applications

Whole-exome sequencing,  
NGS target enrichment, short  
hybridization times

## Products

KAPA HyperPrep Library  
Preparation Kit; KAPA  
HyperExome

# The KAPA HyperCap Workflow v3 yields high-quality results with hybridization times as short as one hour using mechanical DNA fragmentation

*The rapid growth of targeted next-generation sequencing (NGS) applications is increasing the demand for workflows with short turnaround times. The KAPA HyperCap Workflow v3 is a high-performance, streamlined target enrichment solution that includes an overnight (16-20 hours) hybridization step. While this overnight step is standard for hybridization-based workflows, it is a major hurdle in the development of a single-day protocol. This Technical Note describes the development of a short-hybridization target enrichment workflow using the KAPA HyperCap Workflow v3 and KAPA HyperExome Probes. Target-enriched libraries were created using hybridization times as short as 15 minutes, and the sequencing metrics were compared to libraries created using the standard 16-hour workflow. The results show that the sequencing metrics from a 1-hour hybridization are comparable to the 16-hour hybridization workflow, and that hybridization times as short as 15 minutes had only a slight impact on sequencing metrics. The reduction in total workflow time from 22 hours to ~7 hours total may be a worthwhile tradeoff for this slight impact, depending upon the research question.*

## Introduction

The KAPA HyperCap Workflow v3 is a streamlined NGS target enrichment solution that combines high-efficiency KAPA DNA library preparation kits with high-performance KAPA Target Enrichment probes. The workflow accommodates a variety of sample types and starting input amounts into library preparation, and is compatible with mechanical or enzymatic DNA fragmentation using KAPA HyperPrep Kit or KAPA HyperPlus Kit, respectively. Both kits employ KAPA Universal Adapters and Unique Dual-Indexed (UDI) primers, which are designed to ensure high library construction efficiency and low adapter-dimer formation.

The KAPA HyperCap Target Enrichment portfolio includes KAPA HyperExome Probes, a compact, ~43 Mb human whole-exome panel that interrogates exonic regions defined by the CCDS, RefSeq, Ensembl, GENCODE, and ClinVar databases, including medically relevant variants. The HyperCap Workflow v3 takes approximately 22 to 26 hours to complete, from pre-capture library preparation to amplification and clean-up of enriched libraries. The major obstacle in the development of a single-day target enrichment protocol has been the 16- to 20-hour hybridization step, typically performed overnight, during which the target enrichment probes bind to complementary library molecules.

In this study, we tested the feasibility and performance of shorter hybridization times in the KAPA HyperCap Workflow v3 using KAPA HyperExome Probes and the KAPA HyperPrep Library Preparation Kit. *Note: although the results of this pilot study are promising, this short-hybridization protocol is still in development and has not been completely validated, and thus is not fully technically supported by Roche.*

## Methods

### Experimental Design

Libraries were constructed using the KAPA HyperPrep Kit, which utilizes mechanically fragmented DNA. High-quality human gDNA NA12891 (Coriell Institute) was bulk sheared to a mode size of 200 bp using a Covaris ultrasonicator. The fragmented DNA (100 ng) was used as input into automated library preparation, carried out on a Hamilton NGS STAR platform. Libraries were indexed (barcoded) by ligation to KAPA Universal Adapters and subsequent 8 cycles of pre-capture amplification using KAPA Unique-Dual Indexed (UDI) Primer Mixes. The concentration and size distribution of the resulting libraries were determined with the Qubit dsDNA HS Assay kit (Invitrogen) and the Bioanalyzer DNA High Sensitivity Kit (Agilent).

Three replicate target-enriched libraries were prepared for each hybridization time. Each hybridization was performed in a single-plex reaction using 1 µg of library and KAPA HyperExome probes following instructions in the *KAPA HyperCap Workflow v3.0 User Guide*<sup>1</sup> for a capture target size  $\geq 40$  Mb. Hybridization was carried out at 55°C for 15 minutes, 1 hour, 4 hours, or 16 hours (16 hours is recommended in the *KAPA HyperCap Workflow v3.0 User Guide, February 2020 version*, Chapter 5, Step 3, Sub-step 7 of page 24). All other steps were carried out as described.

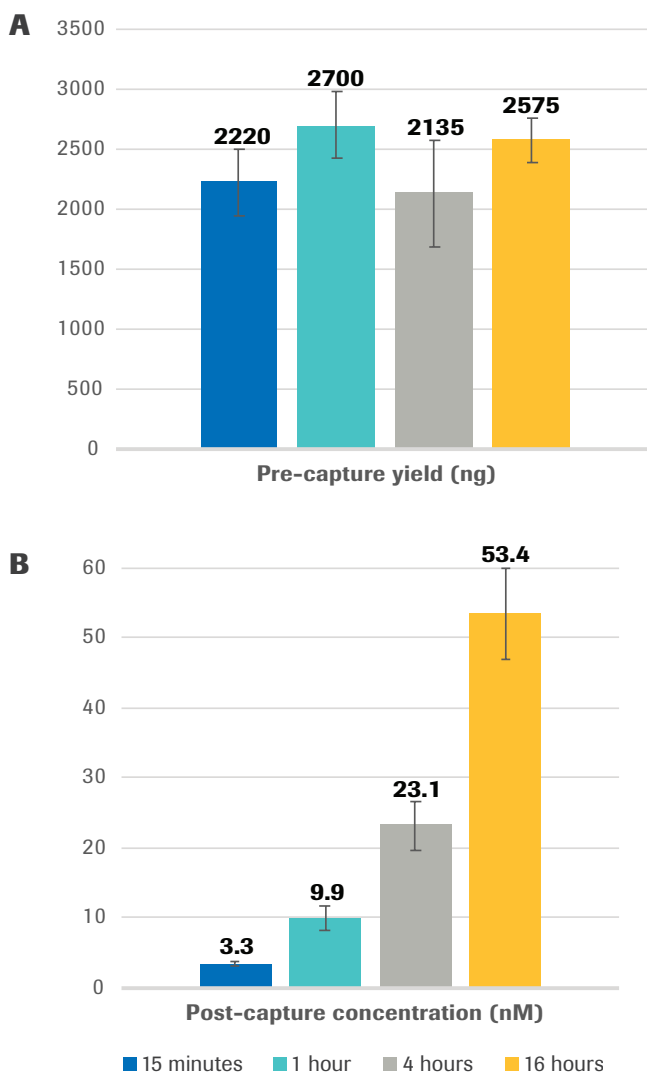
Following bead cleanup, the quantity and quality of amplified enriched libraries were assessed using the Qubit dsDNA HS Assay, the Bioanalyzer DNA High Sensitivity Assay, and the qPCR-based KAPA Library Quantification Kit. Libraries were normalized, pooled, and sequenced on an Illumina® NextSeq® 500 sequencing instrument using the NextSeq High Output kit (2 x 75 bp).

Sequencing data were analyzed according to the technical note *How To... Evaluate NimbleGen SeqCap EZ Target Enrichment Data*.<sup>2</sup> Data was down-sampled to 50X raw coverage. Normalized coverage across GC content was calculated using BEDTools. BEDTools makewindows was used to create 100 bp sliding windows over the length of the capture area. BEDTools nuc and coverage were respectively used to determine GC content and depth of coverage. A combination of R statistical package, Perl and Bash scripts were used to parse the data for graphing purposes.

## Results and Discussion

### Library QC

The quantity and quality of libraries were assessed before and after target enrichment. All pre-capture libraries achieved an average size of 320 bp (data not shown) with a yield  $\geq 1000$  ng (**Figure 1A**), which is the minimum criteria for proceeding to hybridization. All post-capture library yields were sufficient for sequencing (**Figure 1B**).



**Figure 1. All four hybridization times yielded sufficient library for proceeding to sequencing. (A)** Pre-capture yields quantified by Qubit. **(B)** Post-capture concentrations measured by KAPA Library Quantification Kit (qPCR). Bars represent the mean from triplicate libraries and error bars indicate the standard deviation.

### Sequencing metrics

The quality of target-enriched libraries generated with each hybridization time was compared using key sequencing metrics: mean coverage, percent of on-target reads, fold-80 base penalty, percent of duplicate reads, and GC coverage uniformity (**Figure 2**).

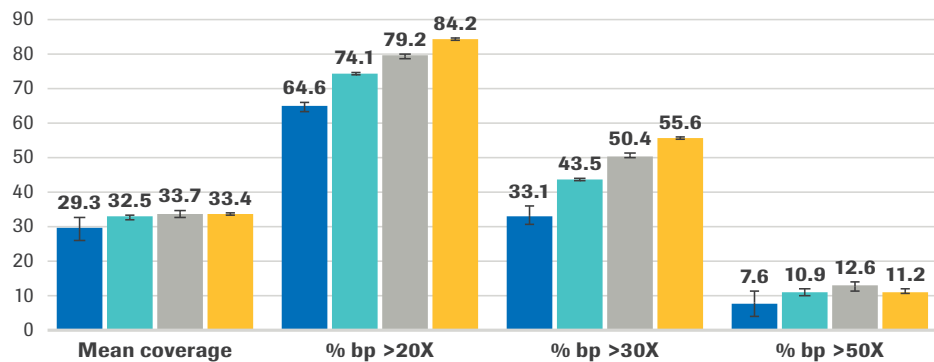
The mean coverage over the entire target region was consistent for all hybridization times. Libraries created with shorter hybridization times displayed a lower percentage of bases covered at 20X and 30X; however, even with a 15-minute hybridization, the percentage of bases covered at 20X was ~65%. At 50X coverage, performance was similar for all libraries.

The overall efficiency of targeted sequencing is described by two sequencing metrics: on-target rate and coverage uniformity (often reported as fold-80 base penalty). The on-target rate refers to the proportion of reads that fall into the target regions, or the genomic regions covered by the panel. All four hybridization conditions tested in this study yielded a similar percent of reads on-target, with a slightly lower rate for libraries hybridized for only 15 minutes (**Figure 3A**).

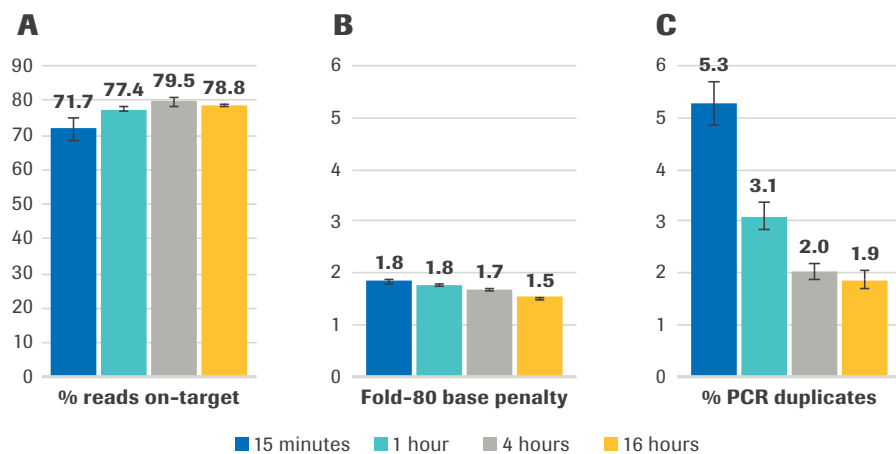
Coverage uniformity describes how evenly the target regions are represented in the sequencing data, and is expressed here as

the fold-80 base penalty. This metric is defined as the fold, or amount, of additional sequencing required to achieve the mean coverage level for 80% of the sequenced bases. A score of 1.0 indicates that no further sequencing is required; lower values close to 1.0 indicate better uniformity. All four hybridization conditions tested in this study exhibited low (below 2.0) fold-80 base penalty scores (**Figure 3B**), with similar scores for all libraries. These results indicate that shorter hybridization times do not substantially impair target enrichment efficiency of the KAPA HyperCap Workflow v3 using KAPA HyperExome.

The level of PCR duplicates in sequencing data indicates library complexity, with low values indicating greater complexity and fewer wasted sequencing reads. In contrast to on-target rate and coverage uniformity, the PCR duplicate rate was affected by reduced hybridization times (**Figure 3C**), reflecting decreased library complexity in shorter protocols. For example, by reducing the hybridization time from 16 hours to 15 minutes, the percent of duplicates increased ~2.8-fold, from 1.9% to 5.3%. However, a ~5% duplication rate is still acceptable for most whole-exome sequencing applications<sup>3</sup>. This result is consistent with the lower post-capture library concentrations observed with samples hybridized for shorter times.

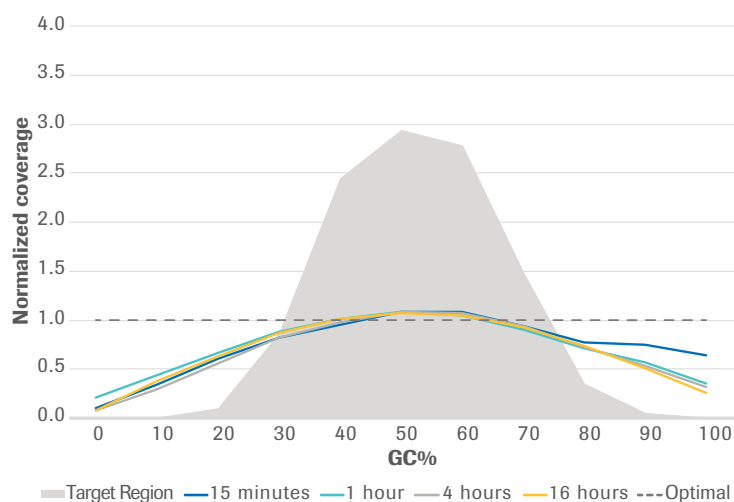


**Figure 2. Shorter hybridization times resulted in reduced sequencing coverage at 20X and 30X coverage, but similar coverage at 50X.** Bars represent the mean from triplicate libraries and error bars indicate the standard deviation.



**Figure 3. Shorter hybridization times have minimal impact on capture efficiency but do affect library complexity.** (A) The percent of on-target reads refers to the percent of mapped, non-duplicate reads overlapping a target region by at least 1 base. (B) Coverage uniformity expressed as fold-80 base penalty. (C) Percent of duplicate reads, associated to the library complexity. Bars represent the mean from triplicate libraries and error bars indicate the standard deviation.

The KAPA HyperExome panel yielded highly uniform enriched libraries with minimal GC% bias regardless of the duration of the hybridization step (**Figure 4**). GC bias describes the relationship between the actual GC content of a region and read coverage across that region; when all regions are equally represented regardless of GC content then the normalized coverage across all regions is 1.0 (**see Figure 4**), and when GC-poor and/or GC-rich regions are under- or over- represented in the sequencing data the normalized coverage is greater or less than 1.0. The normalized coverage of KAPA HyperExome libraries prepared with shorter hybridization times—even as short as 15 minutes—was very similar to the standard 16-hour hybridization over the entire target region.



**Figure 4. Library uniformity across the GC% spectrum is not affected by reduced hybridization duration.** Each curve represents the average of normalized coverage across GC% bins for triplicate libraries. The horizontal line represents the optimal normalized coverage, which would be 1.0 if all sample-to-data steps were completely unbiased. The shaded plot is the GC% distribution of 100 bp windows in the target region.

## Conclusion

Target-enriched NGS workflows with shorter hybridization offer the potential for same-day sample-to-result turnaround times, a valuable time-saver for laboratories that routinely perform targeted NGS. The shorter hybridization method described here enables the generation of sequencing-ready target-enriched libraries in a single workday using the KAPA HyperCap Workflow v3. Target-enriched libraries generated with all four hybridization times tested (15 minutes, 1 hour, 4 hours, and the standard 16 hours) yielded high-quality sequencing metrics, although libraries prepared with shorter hybridization times did demonstrate slightly lower uniformity, reduced coverage, and higher duplicate rates than libraries with 16-hour hybridization. However, the reduction in total workflow time to less than one workday—from 22 hours to 7 hours, with a 1-hour hybridization—may be a worthwhile tradeoff, depending upon the research question. In addition, the performance of libraries hybridized for only 15 minutes suggests that it may be possible to reduce the hybridization time even further.

## References

1. Roche Sequencing Solutions. Instructions for Use. KAPA HyperCap Workflow v3.0. February 2020.
2. Roche Sequencing Solutions. Technical Note. How To Evaluate NimbleGen SeqCap EZ Target Enrichment Data. August 2015.
3. Roche Sequencing Solutions. Application Note. HyperCap: a novel, application-specific approach to streamlined target enrichment. April 2016.

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