



FastGene™ DNA save User Guide

Ambient temperature storage and
transport of purified DNA

Version A

2018





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FOR RESEARCH USE ONLY

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Product Specifications

- 100% recovery of input DNA from 0.05 - 25 µg
- Quality is comparable to input DNA
- Recovery in a volume of 20 - 250 µL
- Increased stability in liquid state for up to 100 hours at room temperature (21-25°C) upon application
- Increased stability in liquid state for up to 8 hours following rehydration of dried DNA, across up to 5 cycles
- Compatible with DNA from cell lines, blood, fresh and frozen tissue, FFPE tissue, cDNA, plasmids, etc., single and double stranded
- Compatible with DNA purified using all standard kits and protocols (Invitrogen, Ambion, QIAGEN)
- Compatible with all common storage buffers, including water, TE, EDTA and citrate
- Use in downstream applications without further purification
 - Does not inhibit PCR, qPCR or expression profiling
- Thermal stability from -80°C to 76°C during transport
 - Exceeds Military specifications for transport (-60°C to 71°C)
 - Exceeds FedEx specifications for transport (-51°C to 60°C)

Storage and Transport

- Store and transport at ambient temperature

Tested storage buffers compatible with

FastGene™ DNA save

- Water, molecular biology grade
- Qiagen Buffer AE
- TE, pH 7.5 and TE pH 8.0 (10mM Tris and 1mM EDTA)
- Low EDTA TE, pH 8.0 (10mM Tris and 0.1mM EDTA)



Simplified Workflow

Apply DNA to FastGene™ DNA save Tubes



Dry for storage or shipping at ambient temperature



To recover, reconstitute in Molecular-grade water for use in downstream applications



DNA is ready for use



FastGene™ DNA save Protocol

DNA Application

1. Apply up to 25 µg of DNA in 20-250 µl to FastGene™ DNA save tube. For concentrated samples add water to a final volume \leq 250 µl.

1. Mix by pipetting up and down 10 times to solubilize the FastGene™ DNA save.

FastGene™ DNA save is supplied as a transparent coating at the bottom of each FastGene™ DNA save Tube.

3. Proceed to Drying Protocol (Page 6).

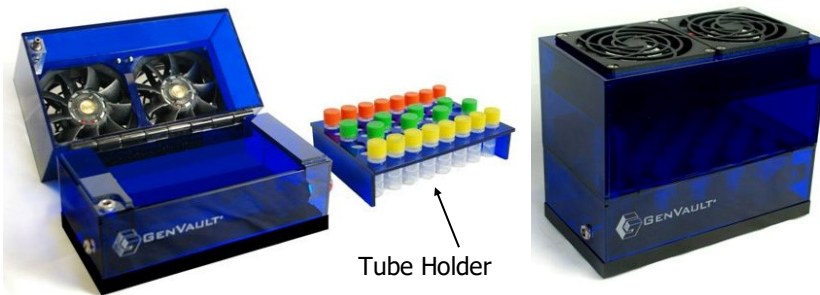
Drying DNA Using a GVGT2001 FastDryer

A FastDryer may be used to dry up to 50 μ L of DNA.

- A GVGT2001 FastDryer may also be used for drying one rack of 0.3ml duster tubes. Please refer to the FastDryer user manual for details at; www.gentegra.com
1. Ensure that the FastDryer is plugged in.
 2. Place unsealed or uncapped tubes or rack in tube/rack holder.
 3. Close the FastDryer lid.
 4. Turn on the FastDryer by pressing the red ON/OFF switch.

Blue lights will illuminate when FastDryer is operating.

5. Dry overnight (16 hours).
 6. Remove samples and cap or seal for storage/transport.
- For details on operation and use of the FastDryer refer to the GenTegra FastDryer User Guide.



FastDryer GVGT2001

FastGene™ DNA save Protocol

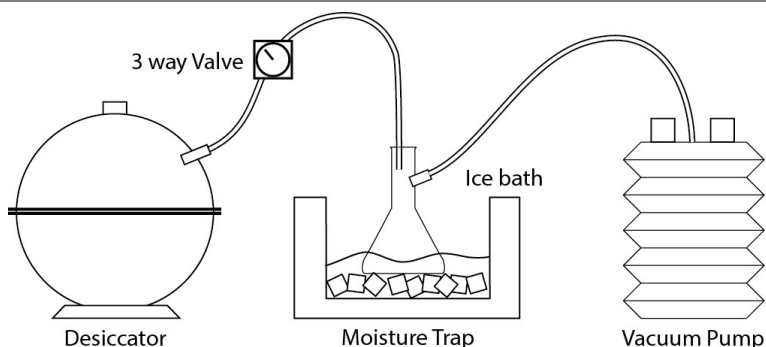
Drying and Storage of DNA

1. Dry DNA according to the methods described in the table below.
 - Drying times will vary depending on application volume.
 - Whatever the drying method, ensure that DNA is completely dry prior to storage.
 - Use SpeedVac on room temperature setting (no additional heat).
 - Drying times for biosafety hood are approximate.

When using 0.5ml screw cap tubes in a FastDryer, volume must be $\leq 50 \mu\text{L}$.

2. When drying is complete, cap or seal tubes/plates and store at room temperature (21-25°C).

Volume	FastDryer™	Vacuum Desiccator or SpeedVac™	Biosafety Hood
$\leq 50 \mu\text{L}$	16 hours	1-4 hours	24 hours
$\leq 100 \mu\text{L}$	32 hours	2-8 hours	48 hours
$\leq 250 \mu\text{L}$	48 hours	4-12 hours	72 hours



FastGene™ DNA save Protocol

DNA Recovery

1. Apply a volume of molecular biology-grade water **equivalent to the input volume**.
2. Mix to solubilize the DNA according to the guidelines in the table below.

Ensure that the final concentration of DNA is **≤200 ng/μl**.

3. Incubate at room temperature (21-25°C) for 15 minutes.
4. Sample is now ready for use.

Product	Recovery Volume and Concentration	Solubilization
0.5 ml Screw Cap Tubes	20-250 μl* ≤200 ng/μl	Cap tubes and vortex for 1 minute

* For sample volumes less than 20 μl see page 17 for special handling information.

Multiple Drying and Rehydration of DNA

Following recovery, an aliquot of DNA may be removed for use, and the sample dried again. This procedure may be repeated five (5) times. This assumes that each rehydration is done at the volume the sample was at the last drying.

For example, a 200 μl sample is applied to a FastGene™ DNA save tube, dried and rehydrated with 200 μl of water. If 50 μl is removed for analysis, leaving 150 μl which is dried again. When this sample is rehydrated it will be done using 150 μl of water. This process can be repeated up to five times with each rehydration volume reduced by the volume of the sample removed. This keeps the concentration of FastGene™ DNA save per μl as in the original sample.

When the remaining volume is 20 μl or less special care should be taken in retrieving the remaining sample.

Product Information

FastGene™ DNA save 0.5mL Screw-cap microtubes	
Catalog #	FG-GTD02-3(Trial), FG-GTD02-100
Tube Volume	0.5 ml
Application Volume	20-250 µl
Application Volume	1-20 µl require special handling
Application Amount	≤ 25 µg
Concentration (DNA application)	Any
Recovery Volume	Equivalent to application volume
Concentration (DNA recovery)	≤200 ng/µl
Drying Method	FastDryer (≤50 µl volume) Vacuum (≤250 µl volume)



Technical Information qPCR

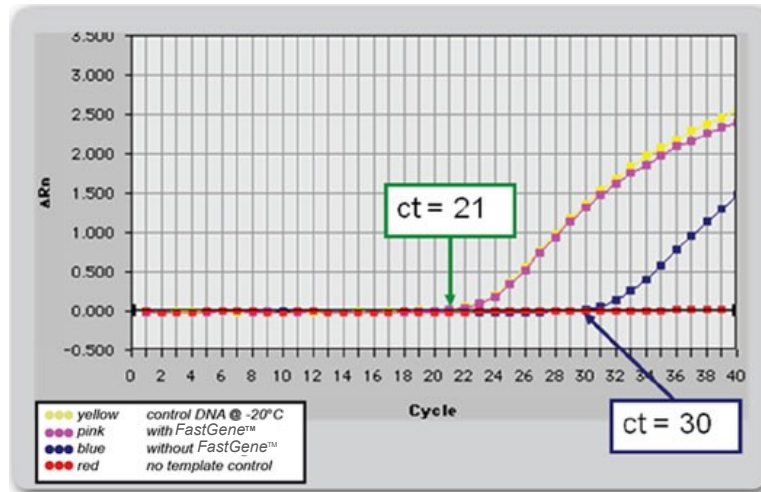


Figure 3. Successful qPCR amplification of DNA stored in FastGene™ DNA save Tubes. Following recovery of DNA after storage at 76°C for two weeks with FastGene™ DNA save, no PCR inhibition was observed even when 26% of the reaction volume was made up of DNA. The green box indicates Ct value of control DNA stored at -20°C and 50ng samples stored at 76°C in the presence of FastGene™ DNA save. The blue box indicates shifted ct values of 50ng samples after storage at 76°C without FastGene™ DNA save.

Long Term Protection and Stability

DNA samples stored on FastGene™ DNA save show no degradation after the equivalent of 16 years storage at ambient temperature. Accelerated stability studies show DNA sample protection with no visible degradation.

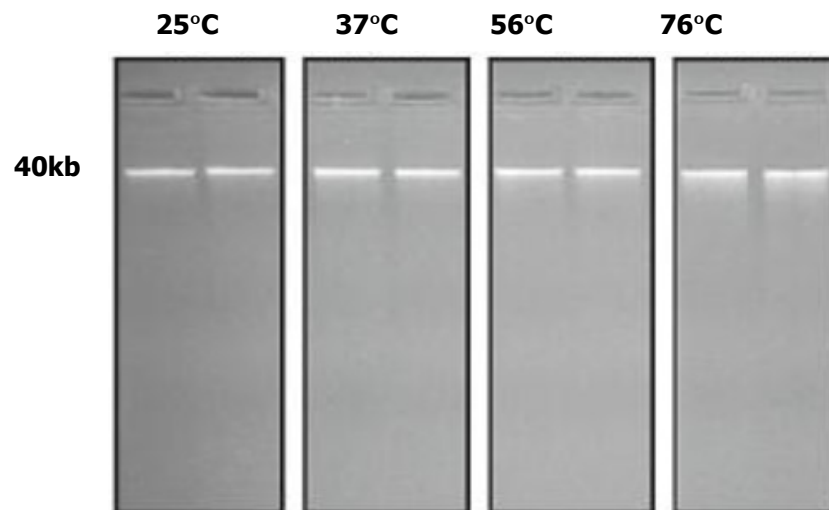


Figure 4. 250 ng/lane genomic DNA stored on FastGene™ DNA save for six months at ambient (25°C) and elevated temperatures.



Technical Information

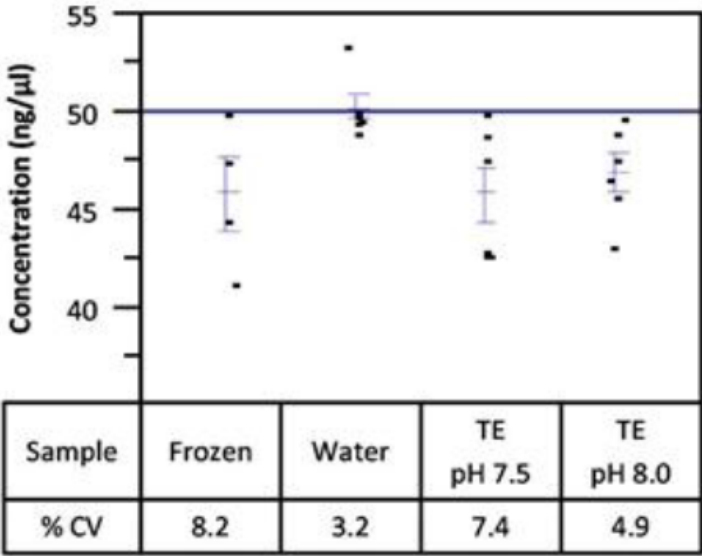


Figure 1. DNA is quantitatively recovered from FastGene™ DNA save Tubes.

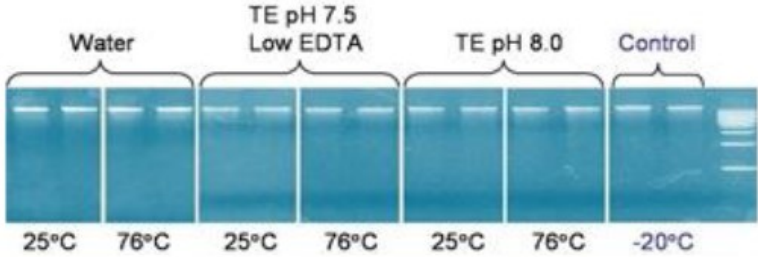


Figure 2. Quality and integrity of DNA stored in FastGene™ DNA save Tubes is identical to DNA stored at -20°C. DNA was stored for 120 days at room temperature (25°C) or 76°C. 120 days of storage at 76°C is equivalent to 10 years of room temperature storage.

Technical Information

Table: Successful genotyping of DNA stored in FastGene™ DNA save Tubes via Illumina and Affymetrix platforms.

		Control (-20°C)	FastGene™ (26°C)
Call Rate	Affymetrix 6.0	99.50%	99.40%
	Infinium IM	99.82%	99.70%
Concordance with frozen control	Affymetrix 6.0		99.80%
	Infinium IM		99.70%

Results using Illumina Infinium IM and Affymetrix 6.0 are identical for DNA stored at -20°C and DNA stored in -20°C and DNA stored in FastGene™ DNA save Tubes at room temperature.



Sample volumes of 20 μ l or less

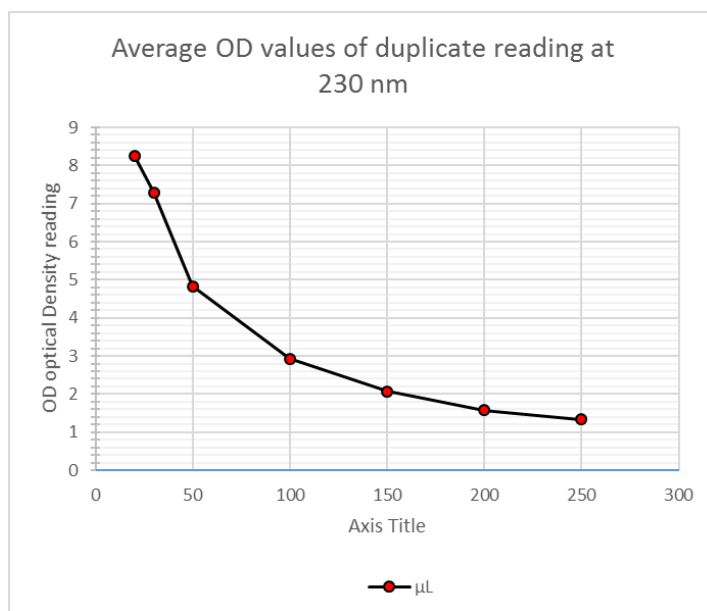
FastGene™ DNA save tubes, microplates and Cluster tubes all start with 21 μ l of FastGene™ DNA save solution being added to the bottom of the tube or well followed by drying. This means the 21 μ l coats the bottom and side walls of each tube/well to the height of 21 μ l. If the sample volume being used is less than 20 μ l it is unlikely that all the FastGene™ DNA save will be dissolved by the sample and these small volumes will make it difficult to wet the sides of the tube to dissolve all the FastGene™ DNA save. For volumes 10 μ l or less this can be an issue.

Small volumes will also tend to stick to the sides of the tube and may not even be in the bottom of the tube when they dry. This means that when the same small volume of water is then used to rehydration the sample it is possible that the rehydration volume may not be in the same place as the original sample. This can lead to apparent sample loss even if the sample is in fact in the tube. Vortexing these small sample volumes can also lead to apparent sample loss as the sample disappears as a coating throughout the inside of the tube. Brief centrifugation may help return the sample to the bottom of the tube but may still lead to losses due to coating of the tube surface.

If small sample volumes are to be used it is recommended that the rehydration volume used be at least 20 μ l to ensure all the original sample is recovered. This dilution of the original sample will not negatively impact the downstream analysis and are likely to improve the actual sample recovery. The 20 μ l low volume cut off is not because FastGene™ DNA save cannot protect small samples but because these small samples are difficult to process conveniently.

Correcting 260/230 ratios

The FastGene™ DNA save chemistry has an absorbance at 230 nm. This absorbance will cause the 260/230 nm ratio values to be different than will normally be expected. The following chart shows the plot of the sample volume vs. OD reading for the FastGene™ DNA save solutions at differing volumes and the table below shows the numerical



µL	OD
250	1.33
200	1.57
150	2.07
100	2.93
50	4.83
30	7.30
20	8.26

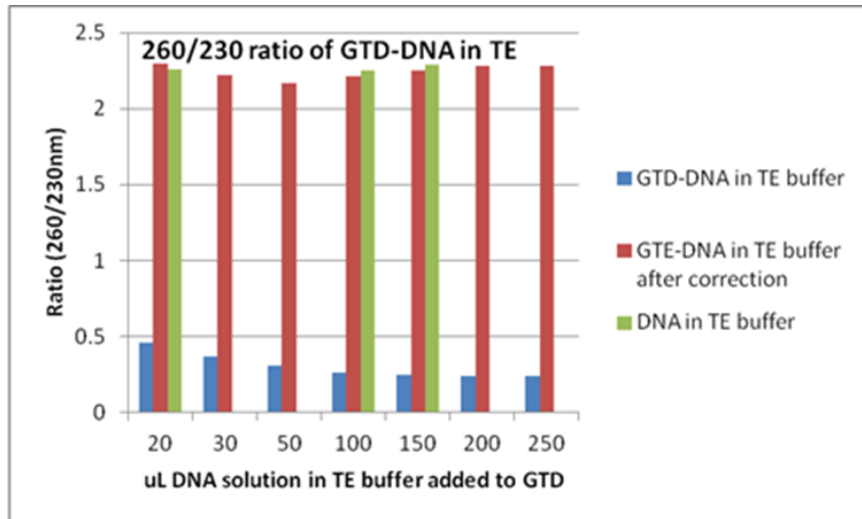
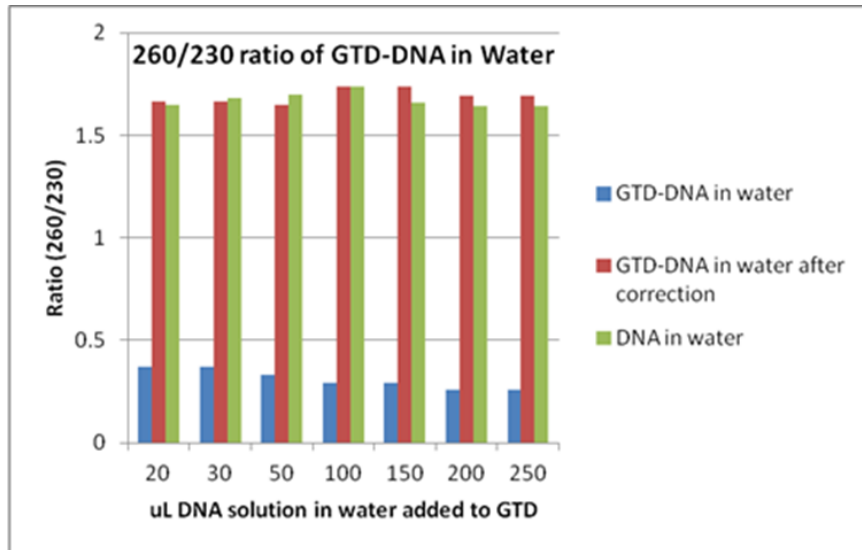
With these values it is possible to create a table of correction values that can be applied to the 260/230 ratios determined using a NanoDrop for example. The absorbance ratio is also affected if TE buffer is being used so a second column is given for the correction factor to use if the DNA & FastGene™ matrix is in TE buffer.

Simply multiply the 260/230 reading you get by the appropriate correction factor.

uL DNA added to GTD	Correction Factor	
	Water	TE
20	4.5	5
30	4.5	6
40	4.5	6
50	5	7
100	6	8.5
150	6	9
200	6.5	9.5
250	6.5	9.5



The following graphs compare 260/230 nm ratios for FastGene™ DNA save plus DNA in water and in TE buffer and the difference with and



Frequently Asked Questions (FAQ)

What is FastGene™ matrix? Is FastGene™ composed of a filter, beads or paper?

FastGene™ DNA save is not a filter, beads or paper; it is an inert chemical matrix.

The FastGene™ DNA save Tubes appear to be empty. Where is the FastGene™ matrix and how can I detect it?

The FastGene™ matrix is supplied as a transparent coating at the bottom of each FastGene™ DNA save Tube. To confirm that the kit you received contains the matrix, simply rehydrate one tube with 35µL of molecular biology grade water and take an absorbance reading at 230nm to detect the FastGene™ matrix.

Can samples stored in low-EDTA TE, water or other buffers be applied to FastGene™ DNA save Tubes?

Yes, refer to Table 2 for a list of storage solutions that are compatible with FastGene™ DNA save Tubes.

What is the maximum concentration of DNA that can be applied to FastGene™ DNA save Tubes?

There is no maximum concentration for application (note that the maximum concentration for recovery is 200 ng/µl) When applying less than 20 µL of DNA, add water to a final volume of ≥20 µl to ensure complete mixing of the DNA with the FastGene™ DNA save. Refer to the tables on pages 14-17 for application volume and mass specifications.

Why is there a minimum recovery volume of 20 µl?

A minimum 20 µl volume is recommended to rehydrate DNA from all surfaces of the tube or well.

Frequently Asked Questions (FAQ) cont'd

Why is there a maximum recovery concentration of 200ng/μL when recovering or concentrating DNA?

Maximum solubility of DNA in water is achieved when the concentration does not exceed 200 ng/μl.

What is the composition of the storage solution after recovery?

After addition of molecular biology water, your samples will be in the same buffer they were stored in at the time of application.

Will the FastGene™ DNA save affect my DNA quantitation? Do I need to blank the spectrophotometer with the FastGene™ DNA save?

The FastGene™ DNA save absorbs at 230 nm. Thus, it will not interfere with readings at Å260 or Å280 and blanking with the FastGene™ DNA save is not required.

How should I store my recovered DNA?

If the recovered DNA is in FastGene™ DNA save we recommend re-drying the DNA solution and storing it at ambient temperature.

Can I use the recovered DNA directly for downstream applications?

Purification is **not** required prior to performing downstream applications. Similar DNA quality is maintained before and after recovery. FastGene™ DNA save does not remove or

inhibit nucleases or other contaminants present in the original sample. When concentrating DNA, please be aware that contaminants will be concentrated along with the DNA.



GenTegra
inside

FastGene™

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